

Laboratory Diagnosis of SARS-Cov-2

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Drexel University College of Medicine Disclosure Statement

Alan T. Evangelista, PhD, D(ABMM)

- No financial or commercial relationships
- No conflicts of interest

Objectives

- Review the principles of PCR (polymerase chain reaction) testing.
- Describe the evaluation and limitations of nucleic acid testing: sensitivity, specificity, positive and negative predictive values.
- Review the diagnostic PCR tests for SARS-CoV-2 comparing viral targets, limit of detection, and specimen types. Describe rapid NAA and rapid Ag tests.
- Discuss availability and specificity of SARS-CoV-2 antibody tests and utility of IgM and IgG results.

Requirements for PCR Test

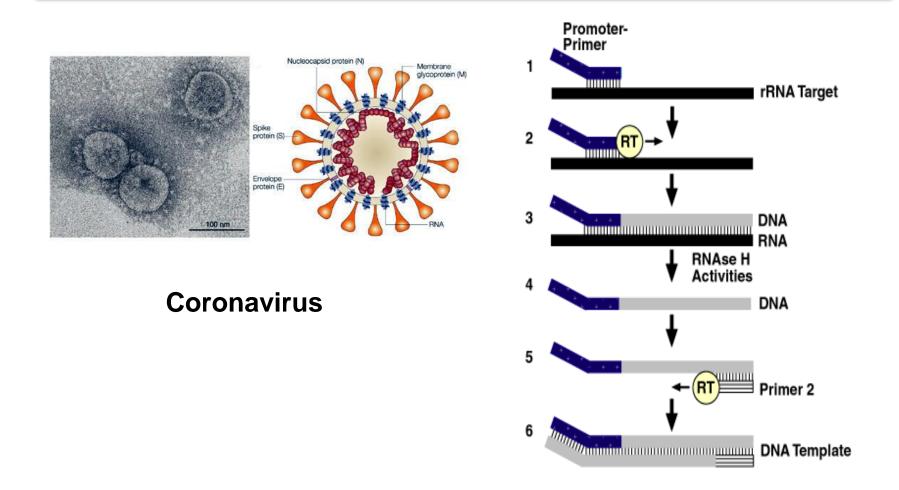
Extraction of specimen: protease +/- beads

Amplification

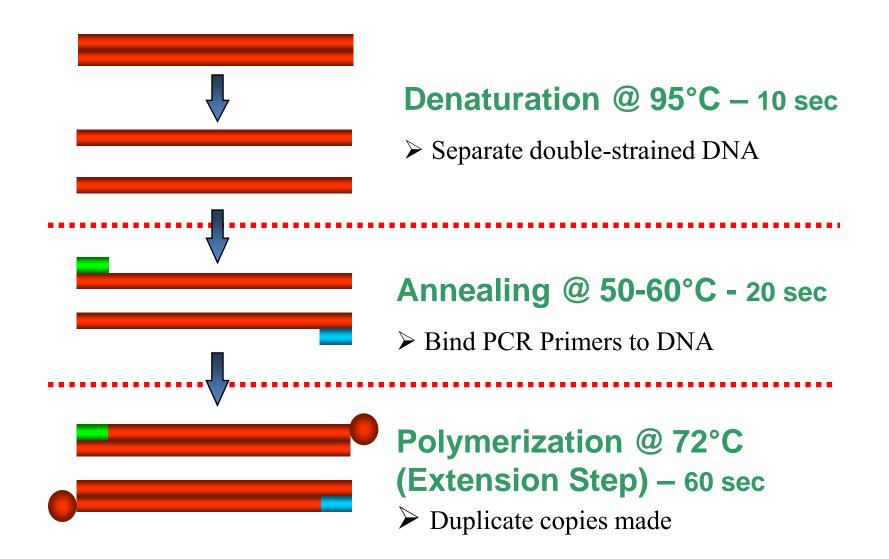
- Identify target sequences: usually 100-10,00 base pairs
- Reverse transcriptase step if RNA virus (make cDNA)
- PCR Primers: a pair of short oligonucleotides (20-30 bp) complementary to opposite strands of DNA flanking the sequence to be detected
- Deoxynucleotides of each base (dNTPs of A,T,C,G)
- Internal control in each sample: QC PCR inhibitors (hemn (hemin) (hemin, lipids)
- Heat stable DNA polymerase enzyme
- Thermal cycler: 95°C -> 55°C -> 72°C

Detection of amplicons: probes w fluorescent reporter

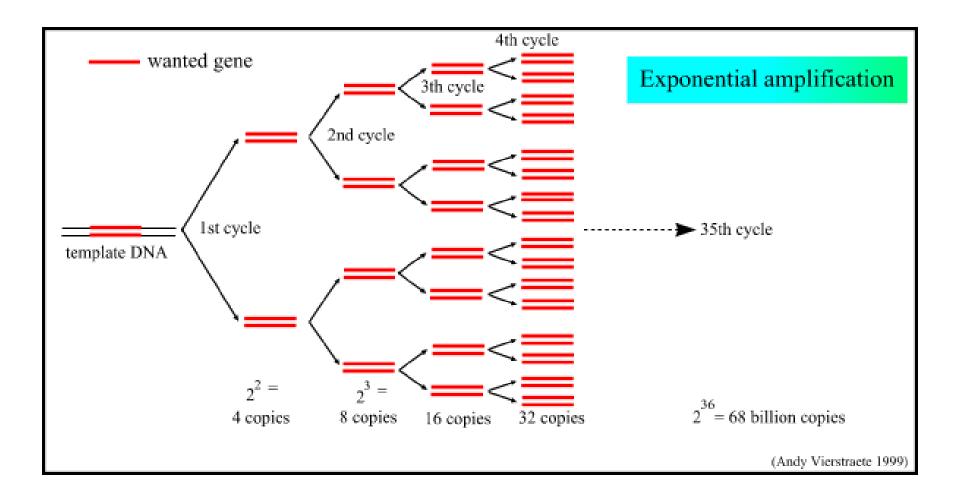
Reverse Transcription PCR (RT-PCR) for RNA viruses



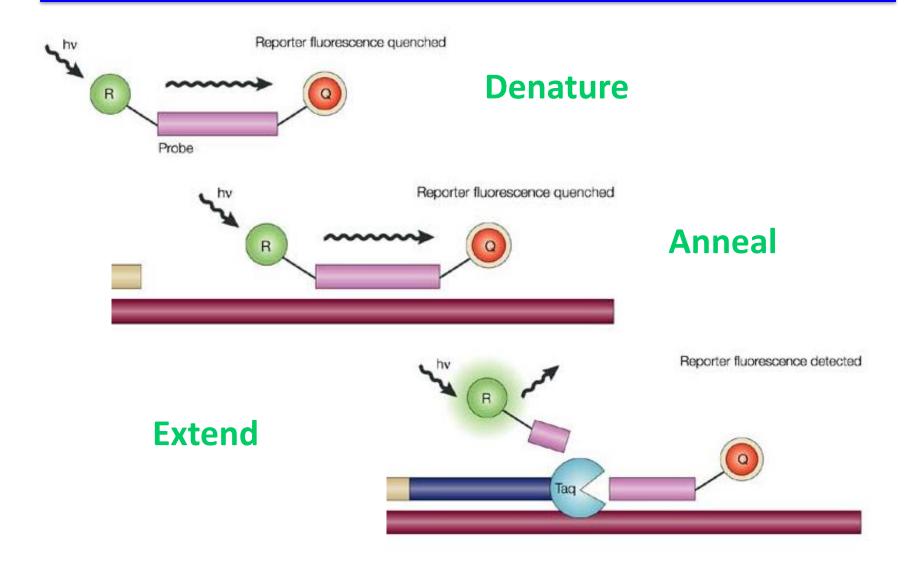
3 Steps of PCR Thermal Cycling



Polymerase Chain Reaction (PCR) Amplification



Fluorescence Detection: Taqman Probes

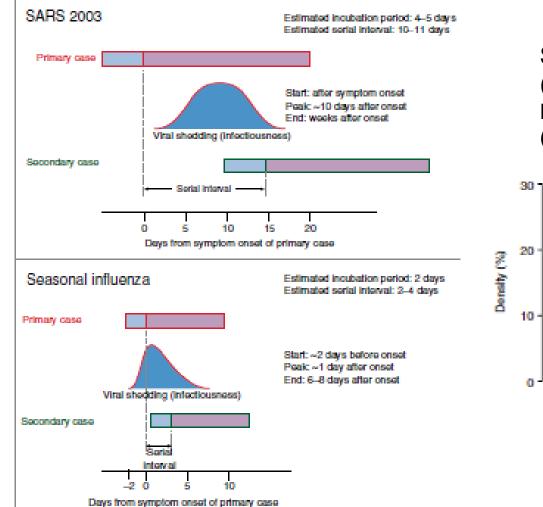


PCR Cycle Threshold Values

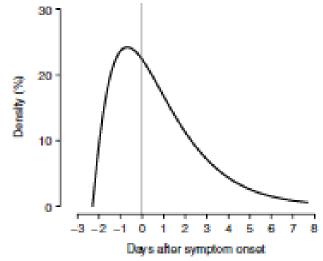
Amplification plot 10. Input n = 81 µg 0.1 µg 10 ng ΔRn 1 ng 1 0.1 ng 10 pg 0.408754 1 pg 0.1 pg NTC 0.1-14 16 18 20 22 24 26 28 30 32 34 36 38 40 2 12 10 6 8 Cycle

New England Biolabs. Luna Probe One-Step RT-qPCR https://www.neb.ca/indexl.php

Temporal Viral Shedding



SARS-CoV-2: virus culture (infectiousness profile) PCR pos up to 14 days after symp (some residual RNA up to 21 days)



He, X, et al. Temporal dynamics in viral shedding and transmissibility of COVID-19. Nat Med 15 Apr 2020. https://doi.org/10.1038/s41591-020-0869-5

SARS-CoV-2 Viral Culture vs PCR Cycle Threshold Values

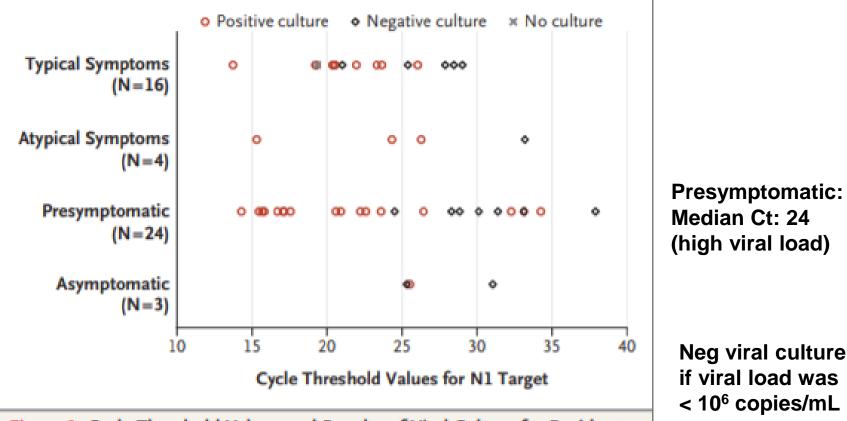
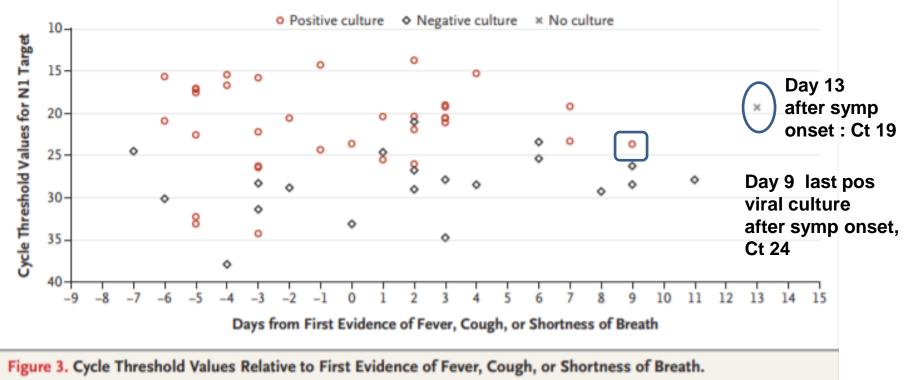


Figure 2. Cycle Threshold Values and Results of Viral Culture for Residents with Positive SARS-CoV-2 Tests According to Their Symptom Status.

Shown are N1 target cycle threshold values and viral culture results for 47

Arons, M, et al. Presymptomatic SARS-CoV-2 infections and transmission in a skilled nursing facility. NEJM 24 Apr 2020. doi: 10.1056/NEJMMoa2008457

Temporal SARS-CoV-2 Ct Values from Symptomatic Patients



Shown are N1 target cycle threshold values and viral culture results for each resident's positive tests for SARS-CoV-2

Arons, M, et al. Presymptomatic SARS-CoV-2 infections and transmission in a skilled nursing facility. NEJM 24 Apr 2020. doi: 10.1056/NEJMMoa2008457

Test Sensitivity and Specificity

• Sensitivity:

- Ability of the test to correctly identify those patients with the disease
- A test with 90% sensitivity: 10% with disease go undetected (10% false negatives)
- Analytic sensitivity of PCR: depends of limit of detection
- Clinical sensitivity of PCR: depends on specimen collection and time of collection
- Specificity:
 - Ability of the test to correctly identify those patients without the disease
 - A test with 90% specificity: 10% without the disease are incorrectly identified as test positive (10% false positives)

Disease Prevalence of 5%

- Test Sensitivity and Specificity of 95%
 - Negative predictive value: 99.7%
 - False neg rate: 0.3%
 - Positive predictive value: 50%
 - False pos rate 50%
- Test Sensitivity and Specificity of 99%
 - Negative predictive value: 99.9%
 - False neg rate: 0.1%
 - Positive predictive value: 84%
 - False pos rate 16%

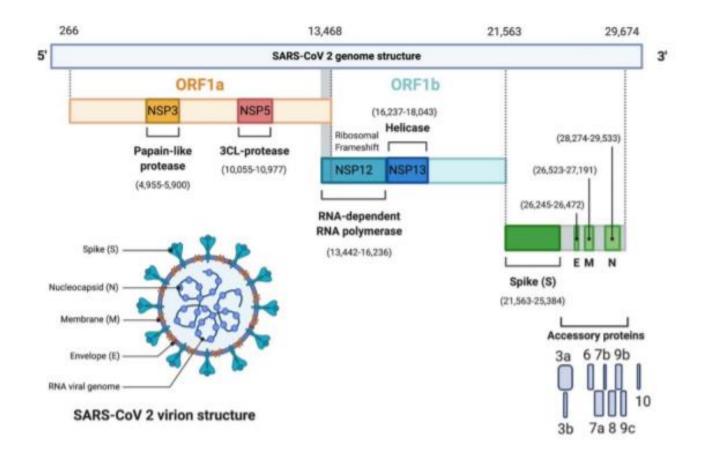
Cairns, E. COVID -19 antibody tests face a very specific problem. Evaluate Vantage COVID-19 Report.16 Apr 2020. www.evaluate.com

Specimen Source Sensitivity for SARS-CoV-2 PCR Assays (Symptomatic Patients)

Specimen	Sensitivity	Collection	Swab Type
Nasopharyngeal (NP)	97%	HCW	Flocked, Synthetic
Nasal (mid-turbinate)	100%	HCW Self-collection	Flocked, foam, synthetic
Saliva	85%	Self-collection	Wide tube, Urine cup
Nasal (anterior nares)	76%	HCW Self-collection	Flocked, foam, synthetic
Oral (throat)	56%	HCW Self-collection	Synthetic

Hanson, et al. IDSA Guidelines Diagnosis of COVID-19. 6 May 2020 .www.idsociety.org/COVID19guidelines/dx

SARS-CoV-2 RT-PCR Targets



Alanagreh, et al. Human coronavirus disease COVID-19.Pathogens 9:331 MDPI 29 Apr 2020. doi:10.3390/pathogens9050331

RT-PCR Assays for SARS-CoV-2 (96 well, high throughput)

Company	Instrument Name	PCR Kit Sample size	Targets	Limit of Detection	Assay Time
CDC	ABI 7500 Fast DX	TaqPath COVID-19 (110 μL sample) (5 μL eluate extract)	N1 N2	150- 780 copies/mL	4-5 hr
Roche	Cobas 6800	Cobas SARS-CoV-2 (400 μL sample)	Orf 1ab E	150 copies/ mL	3-4 hr
Abbott	m2000	Alinity m SARS-CoV-2 (100 μl sam, 5 μl ext)	N RdRp	150 copies/mL	4-5 hr
Thermo Fisher	ABI 7500 Fast DX	TaqPath CovID-19 (400 μl sample) (5 μl extract)	N S Orf 1ab	250 copies/mL	4 hr
LabCorp	ABI Quant Studio Flex	TaqPath COVID-19 (200 μL s)	N1 N2	150 copies/mL	3-4 hr
Quest	ABI 7500 Fast DX	TaqPath COVID-19 (250 μL s) (10 μL ext)	N1 N3	150 copies/mL	4 hr

Commercial RT-PCR Assays for SARS-CoV-2

Company	Instrument Name			Limit of Detection	Assay Time
Cepheid	GeneXpert Xpress (300 µL sample)	Single cartridge 12,16,24/ins	N2 E	200 copies/mL	45 min
DiaSorin Molecular	Liaison MDX (50 µL sample)	8 test disc	S Orf 1ab	40ª-150 copies/mL	80 min
GenMark	ePlex (200 µL sample)	Single cartridge 3-24/instr	Ν	1,000 copies/mL ^a	3-4 hr
Biofire	Torch Film Array (300 μL sample)	Single pouch (22 targets) 8-12/instr	S M	160 copies/mL	45 min
Luminex	Aries (200 μL sample)	Single cassette 6-12/instr	N Orf 1ab	75,000 copies/mL	2 hr

a: Zhen, et al. Comparison of 4 molecular in vitro assays for SARS-Cov-2. Medrxiv https://doi.org.1101/2020.04.17.20069864

Commercial RT-PCR Assays for SARS-CoV-2

Company	Instrument Name	Format	Targets	Limit of Detection	Assay Time
Becton Dickinson	BD Max (750 μL sample)	Single cartridge 12,16,24/instr	N1 N2	200 copies/mL	2 hr
Hologic	Panther Fusion (500 µL sample)	10 test cartridge 120/instr	Orf 1a Orf 1b	83-625 copies/mL ^a	3 hr

a: Zhen, et al. Comparison of 4 molecular in vitro assays for SARS-Cov-2. Medrxiv https://doi.org.1101/2020.04.17.20069864

Commercial NAA Assays for SARS-CoV-2

Company	Instrument Name	Format	Targets	Limit of Detection	Assay Time
Abbott	ID NOW Direct-dry	Single cartridge	RpRp (template)	20,000 copies/mL ^a	15 min
	ID NOW in VTM	Single cartridge	RpRp	15-30% false neg ^a	15 min

Rapid Commercial Ag Assays for SARS-CoV-2

Company	Instrument Name	Format	Targets	Limit of Detection	Assay Time
Quidel	Sofia: lateral Flow FIA (120 µl sample)	Single test cassette	N (acute infection)	850 copies/mL (confirm neg with molec test) IFU 80% sensitivity	15 min

a: Basu, et al. Performance of NAA by Abbott ID NOW COVID-19. Medrxiv <u>https://doi.org.1101/2020.05.11.089896</u> (posted 12 May 2020)

Commercial Ab Assays for SARS-CoV-2

Company – Test	Instrument Name	Format	Targets	Sensitivity Specificity	hCoV -X PPV at 5% prev	Assay Time
DiaSorin IgG	Liaison XL (Chemilum)	12/rack 120/run	S1 S2	97.6% 99.3%	No 88%	30 min
	Pos Percent Agreement	<u><</u> 5 days 6-14 days <u>></u> 15 days	25% 90% 97.6%			
Abbott IgG	Architect (Chemilum)	5/rack 50/run	Ν	100% 99.0%	Yes 92.9%	30 min
Eurolmmun IgG	ELISA: (man/auto)	96 well	S1	90% 99.1%	No 88%	2-3 hr
Ortho Clinical Diag IgG & Total	Vitros (Chemilum)	10/rack 100/run	S1	87.5% 99.1%	No 88%	1 hr

https://www.fda.gov/medical-devices/emergency-situations

Summary of SARS-CoV-2 Diagnostic Tests

I. Acute Infection

• Molecular testing: RT-PCR, NAA less sensitive

II. Recent Infection

- IgM: 1-7 days after symp, 85% of patients (Guo, CID 2020)
 - IgM useful for suspected COVID patients w neg molec PCR
 - IgM is not a viral neutralizing Ab
- IgA: 1-3 days after symp onset
 - IgA most abundant Ig in mucosal surfaces
 - IgA has viral neutralizing activity

III. Recent or Remote Past Infection

- IgG: 6-14 days after symp onset, 90% of patients
 - IgG has viral neutralizing activity
 - IgG is a long lasting Ab
 - Low level false pos rate due to autoantibodies

Utility of SARS-CoV-2 IgG Tests

- I. Screening of Recovered COVID-19 Patients
 - Convalescent plasma: treat acutely ill patients
 - Donor plasma with Ab titer of 1:160
- **II. SARS-CoV-2 Seroprevalence studies**
 - Prevalence of total disease in community
 - Asymptomatic prevalence 50%, mild infection 30%
 - Screen HCW for immune status & patient exposure
 - Guidance for return to work status (PCR + Ab)

III. Monitor Immune Responses for Vaccine Candidates

- Prescreen individuals prior to vaccine clinical trial
- Monitor temporal immune response
- Determine if serologic assay is able to detect neutralizing Ab
- Determine duration of protective immunity

Theel, et al. The role of antibody testing for SARS-CoV-2: Is there one?. JCM doi:10.1128/JCM.00797-20 (posted 29 Apr 2020)